Contents lists available at ScienceDirect

Cancer Letters



journal homepage: www.elsevier.com/locate/canlet

Advances in the epidemiology of pancreatic cancer: Trends, risk factors, screening, and prognosis

Jie Cai^{a,1}, Hongda Chen^{b,1}, Ming Lu^b, Yuhan Zhang^b, Bin Lu^b, Lei You^a, Taiping Zhang^a, Min Dai^{b,*}, Yupei Zhao^{a,**}

^a Department of General Surgery, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, 100730, China

^b Office of Cancer Screening, National Cancer Center/National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences, Peking Union Medical College, Beijing, 100021, China

ARTICLE INFO

Keywords: Pancreatic cancer Trend Screening Prediction model Biomarker

ABSTRACT

Pancreatic cancer is a malignancy with poor prognosis and high mortality. The recent increase in pancreatic cancer incidence and mortality has resulted in an increased number of studies on its epidemiology. This comprehensive and systematic literature review summarizes the advances in the epidemiology of pancreatic cancer, including its epidemiological trends, risk factors, risk prediction models, screening modalities, and prognosis. The risk factors for pancreatic cancers can be categorized as those related to individual characteristics, lifestyle and environment, and disease status. Several prediction models for pancreatic cancer have been developed in populations with new-onset diabetes or a family history of pancreatic cancer; however, these models require further validation. Despite recent progress in pancreatic cancer screening, the quantity and quality of related studies are also unsatisfactory, especially with respect to the identification of high-risk populations and development of effective screening modality. Apart from the populations with familial genetic risk and those at a high risk of sporadic pancreatic cancer, risk factors such as new-onset diabetes may be a new direction for timely intervention. We hope this work will provide new ideas for further prevention and treatment of pancreatic cancer.

1. Introduction

Pancreatic cancer, a common malignant tumor, which frequently occurs as pancreatic adenocarcinoma, is characterized by poor prognosis, with an overall 5-year relative survival rate of approximately 10% [1]. Pancreatic neuroendocrine tumor, accounting for less than 5% of pancreatic cancer, has different characteristics and treatment methods compared to pancreatic adenocarcinoma [2], and is not the focus of the present review. Surgical resection at an early stage is currently the only effective treatment. Therefore, early diagnosis and timely surgical intervention are currently the only effective means to improve outcomes in pancreatic cancer patients. It is important to identify the related risk factors and populations that are at a high risk of pancreatic cancer, in addition to achieving early diagnosis based on clinical examinations or biomarkers. Recent decades have witnessed a series of advances in

¹ These authors contributed equally to this study.

https://doi.org/10.1016/j.canlet.2021.06.027

Received 26 March 2021; Received in revised form 9 June 2021; Accepted 25 June 2021 Available online 30 June 2021 0304-3835/© 2021 Elsevier B.V. All rights reserved.

epidemiological and clinical research on pancreatic cancer [3–10]. This review summarizes the advances in research on pancreatic cancer in terms of its epidemiological trends, risk factors, risk prediction models, screening modalities, and prognosis, which may provide important references for the further prevention and treatment of pancreatic cancer.

2. Epidemiology of pancreatic cancer

According to the Global Cancer Observatory (GLOBOCAN) 2020 [11], an estimated 495,773 patients were newly diagnosed with pancreatic cancer in 2020 worldwide, ranking pancreatic cancer 12th among all malignant tumors. The global crude incidence rate was $6.4/10^5$, and the age-standardized incidence rate (ASR) was $4.9/10^5$. An estimated 466,003 deaths were attributed to pancreatic cancer in 2020, resulting in pancreatic cancer ranking 7th among all malignant tumors,





^{*} Corresponding author.

^{**} Corresponding author.

E-mail addresses: daimin2002@hotmail.com (M. Dai), zhao8028@263.net (Y. Zhao).

with a crude mortality of $6.0/10^5$ and an ASR of $4.5/10^5$. The incidence and mortality (both crude and ASR) were higher in men than in women. Asia contributed the most to both the new diagnosed cases (47.1%) and cancer-related deaths (48.1%) worldwide. The incidence and mortality rates of different countries worldwide in 2020 are shown in Fig. 1.

An analysis of recent trends in pancreatic cancer in 48 countries by Huang et al. [7] showed significant increases in the incidence of pancreatic cancer in men in 14 countries (average annual percent changes [AAPCs], 8.85–0.41) and in women in 17 countries (AAPCs, 6.04–0.87). Similarly, increased mortality was observed in men in 8 countries (AAPCs, 4.20–0.55) and in women in 14 (AAPCs, 5.83–0.78) countries. An increase was also observed in individuals older than 50 years of age in 18 countries. Malta showed the fastest-growing incidence in women, with an AAPC of 6.04%. Iceland reported the most drastic increase in men, with an AAPC of 8.85%. In terms of mortality, the Philippines showed the most significant increase in both sexes, with AAPCs of 4.20% in men and 5.53% in women [7].

The human development index (HDI), calculated according to life expectancy, education level, and quality of life, is used to evaluate the state of national development. A higher HDI was associated with an increased incidence and mortality of pancreatic cancer [7]. In the US, pancreatic cancer is the 4th leading cause of death among all cancers [12] and is predicted to be the second leading cause of death by 2030 [13]. In EU countries, the incidence of pancreatic cancer ranks 8th among all malignant tumors, and the cancer-related death rate ranks 6th. The total incidence of pancreatic cancer is expected to increase by an estimated 30% by 2040 [14]. In China, there were an estimated 95, 000 (crude incidence rate: $6.92/10^5$) and 85,000 (crude mortality rate: $6.16/10^5$) new cases and deaths, respectively, due to pancreatic cancer in 2015, ranking 10th and 6th among all malignant tumors in China, respectively [15].

Pancreatic cancer imposes a great burden on human health globally. There was a 2.1-fold increase in global disability-adjusted life years (DALYs) due to pancreatic cancer from 1990 to 2017, increasing from 4.4 million (95% confidence interval [CI]:4.3–4.5) to 9.1 million (95% CI, 8.9–9.3) [16]. In 2017, the DALY of pancreatic cancer in China was 1.89 million (DALY rate: $133.87/10^5$), approximately three times the disease burden in 1990 [17].



Fig. 1. Estimated age-standardized incidence rates (World) for pancreatic cancer in 2020, both sexes (the figure was mapped using published data from GLOBO-CAN 2020).

3. Risk factors for pancreatic cancer

In general, the risk factors for pancreatic cancer can be categorized as those related to individual characteristics, lifestyle and environment, and disease status. Among them, the risk factors associated with individual characteristics mainly include age, sex, race, ABO blood group, family history, and genetic mutations. The risk factors related to lifestyle and environment include dietary habits, exposure to trace elements, smoking, drinking, physical activity, and obesity. The disease status aspect includes chronic pancreatitis, diabetes, infection, etc [18]. An overview of the risk and preventive factors for pancreatic cancer is shown in Table 1.

3.1. Individual characteristics

3.1.1. Age

Pancreatic cancer mainly occurs in elderly individuals. In the US, most patients are diagnosed with pancreatic cancer in later life, with a median age of 70 years at diagnosis, with only 10.6% of diagnoses being made in patients before the age of 55 years [1]. In China, both the incidence (ASR: $5.02/10^5$ in 2017) and mortality (ASR: $5.67/10^5$ in 2017) of pancreatic cancer were the highest in the population aged 85–90 years. The DALY was the highest in people aged 70–74 years in both 1990 and 2017 [17]. Pancreatic cancer may be associated with age-related pancreatic morphological and pathological changes such as telomere dysfunction [19].

3.1.2. Sex

The incidence of pancreatic cancer is generally higher in men than in women. The reported global incidence (ASR) in men was $5.7/10^5$ and $4.1/10^5$ in women. In Polynesia, the incidence (ASR) in men was approximately 3.6 times that in women (7.9/2.2). However, there were no significant sex differences in terms of mortality (ASR) [11]. In China, the incidence in men (crude: $7.67/10^5$; ASR: $5.06/10^5$) was also slightly higher than that in women (crude: $6.14/10^5$; ASR: $3.54/10^5$) [15].

Table 1

Factors	Risk	References
Individual characteristics		
Age (older)	+++	[1,17]
Gene mutations	+++	[5,10,29,36]
Race (black)	++	[1,21,22]
Family history	+	[28]
Gender (male)	+	[11,15]
Blood group (A/B/AB)	+	[23-26]
Lifestyle and environment		
Exposure to iron, cadmium, arsenic and lead	++	[77-80]
High consumption of red and processed meat	+	[41,42]
Sugar-sweetened foods and drinks	+	[43-45]
Excessive alcohol consumption	+	[61,62]
Smoking	+	[64,65]
Obesity	+	[69,71]
Soy products	(+)	[53]
Exposure to selenium and nickel	(-)	[80]
Coffee	(-)	[47-50]
Vitamin D	(-)	[58,59]
Vegetables, fruits, nuts, whole grains	-	[54–57]
Physical activity	-	[66,67]
Aspirin use	-	[73,74]
Disease status		
Chronic pancreatitis	+++	[82]
Type II diabetes	++	[3]
Hepatitis B virus infection	++	[99]
Pancreatic cystic lesions	+	[86–94]
Helicobacter pylori infection	+	[100]

 \dagger : +++= very strong risk increase (>3-fold) . ++ = strong risk increase (2~3-fold). + =moderate risk increase (1~2-fold).

- = risk reduction. Parentheses show probable but not fully established associations.

However, studies have not shown the protective effect of estrogen against pancreatic cancer [20]. Therefore, the causes and mechanisms of sex differences, which may be related to genetic and lifestyle differences between men and women, require further exploration.

3.1.3. Race and ethnicity

The risk of pancreatic cancer varies among ethnicities. According to the Surveillance, Epidemiology, and End Results (SEER) database (2013–2017), the ASR in males was the highest in black ($(16.9/10^5)$), followed by non-Hispanic ($(15.2/10^5)$) and white ($(15.0/10^5)$) populations, and the lowest in Asian/Pacific Islanders ($(11.0/10^5)$). In women, the highest incidence was also observed in the black ($(14.1/10^5)$), followed by non-Hispanic ($(11.7/10^5)$) and white ($(11.6/10^5)$) populations, while the lowest incidence was observed in American Indian/Alaska natives ($7.8/10^5$) [1]. Excess risk of pancreatic cancer among American black populations may be attributed to race-based differences in the metabolism of cigarette smoke, higher levels of cigarette smoking, high-calorie diets, heavy alcohol consumption, obesity, long-standing diabetes, and low-income level [21]. In addition, pancreatic cancer appears to be less aggressive and more survivable in Asian patients residing in the US than in non-Asians residing in the US [22].

3.1.4. Blood group

Individuals with blood types A, B, and AB are at a higher risk than those with blood type O. Compared to blood type O, the odds ratios (ORs) for pancreatic cancer in individuals with types A, AB, and B were 1.38 (95% CI: 1.18–1.62), 1.47 (95% CI, 1.07–2.02), and 1.53 (95% CI, 1.21–1.92), respectively [23]. This difference may be related to the single nucleotide polymorphism of ABO, rs505922, which causes a strong linkage disequilibrium in the O/non-O blood group alleles [24, 25]. A pooled analysis of 24 studies on the association between ABO blood type and pancreatic cancer among cytotoxin-associated gene A (CagA)-endemic and CagA-nonendemic individuals showed that compared to group O, individuals with group A blood type in both CagA-nonendemic and CagA-endemic populations showed increased risks (OR_{pooled}, 1.40; 95% CI, 1.32–1.49). Meanwhile, group B (OR 1.38; 95% CI, 1.16–1.64) and AB (OR, 1.52; 95% CI, 1.24–1.85) were associated with higher risk only in non-endemic populations [26].

3.1.5. Family history and gene mutations

In addition, pancreatic cancer also shows familial characteristics. Approximately 5–10% of pancreatic cancer patients have a family history of pancreatic cancer [27]. A meta-analysis showed that people with a family history have a significantly increased risk of pancreatic cancer, with a relative risk (RR) of 1.80 (95% CI, 1.48-2.12) [28]. Gene mutations, including those of germ and somatic cells, and genetic syndrome are also associated with pancreatic cancer. Hereditary pancreatic cancer can present in the context of several hereditary syndromes, including Peutz-Jeghers syndrome (PJS), hereditary pancreatitis (HP), familial atypical multiple mole melanoma (FAMMM), hereditary breast and ovarian cancer syndrome (HBOC), Lynch syndrome (LS), and familial adenomatous polyposis (FAP) [29]. The most frequent genetic alterations are those in breast cancer gene 2 (BRCA2), partner and localizer of BRCA2 (PALB2), ataxia-telangiectasia-mutated (ATM), and CDKN2A/p16, and, less frequently, BRCA1, adenomatous polyposis coli (APC), MutL homolog 1 (MLH1), MutS homolog 2 (MSH2), MSH6, PMS1 homolog 2 (PMS2), serine protease 1 (PRSS1), and serine/threonine kinase 11 (STK11) [29].

Most of these hereditary syndromes associated with pancreatic cancer are inherited autosomal dominant gene mutations. PJS, with a mutation in *STK11* (also known as *LKB1*), is characterized by the presence of hamartomatous polyps on the gastrointestinal tract or mucosae. A meta-analysis showed a cumulative lifetime risk of pancreatic cancer of 36% in patients with PJS [30]. HP is characterized by chronic pancreatitis and recurrent acute pancreatitis, the causative genes of which include *PRSS1*, *SPINK1*, *PRSS2*, and chymotrypsin C (*CTRC*). The

cumulative risk of pancreatic cancer in patients with HP varies between 7.2% [31] and 53.5% [32]. FAMMM, also an autosomal dominant inherited syndrome but with incomplete penetrance, is characterized by multiple atypical nevi progressing to melanoma. Approximately 30%-40% cases of FAMMM syndrome are caused by germline mutations in p16/CDKN2A [33], and the cumulative risk of pancreatic cancer is as high as 17% [34]. HBOC is genetically caused by inactivating mutations in BRCA1, BRCA2, and PALB2. The reported mutation rates of these three gene were 0.59%, 1.95%, 0.40%, respectively [35]. BRCA2 mutation carriers had a 21.7-fold increased risk of pancreatic cancer, compared to a 2- and 6-fold increase in risk of pancreatic cancer in BRCA1 and PALB2 mutation carriers, respectively [29,36]. LS, associated mainly with an increased risk of colorectal cancer, develops from germline mutations in mismatch repair genes (MLH1, MSH2, MSH6, and PMS2), with reported mutation rates of 0.5% in patients with pancreatic cancer [35]. Recent evidence has indicated an association between pancreatic cancer and LS mainly in patients carrying MLH1 mutations, who showed an 8.6-fold increase in the risk of pancreatic cancer and a cumulative risk of 3.7% of developing pancreatic cancer 70 years of age [5,10]. FAP is associated with germline mutations in APC, with a cumulative risk of pancreatic cancer of 1.7% at 80 years of age [37]. Patients with other hereditary syndromes, such as Li-Fraumeni syndrome (LFS), associated with TP53 mutations; ataxia telangiectasia (AT), associated ATM mutations; and cystic fibrosis (CF), associated with CFTR mutations, showed <5% lifetime cumulative risk of pancreatic cancer [38].

3.2. Lifestyle and environment

Lifestyle and environment are modifiable risk factors and important foci of disease prevention and health promotion strategies. A healthy lifestyle is an effective method for pancreatic cancer prevention and control [39]. As an important digestive organ, the pancreas participates in the digestion and metabolism of sugars, proteins, and lipids. Thus, diet plays an important role in the pathogenesis of pancreatic cancer.

3.2.1. Red meat and saturated fat

Red meat and saturated fat are proven risk factors for gastrointestinal tumors, such as colorectal cancer [40]. Moreover, excessive intake of red meat and saturated fat may also increase the risk of pancreatic cancer [41]. The "Diet, nutrition, physical activity, and pancreatic cancer" revised in 2018 [42] reported a non-significant association between per 100 g/d increases in red meat consumption and the risk of pancreatic cancer (RR, 1.14; 95% CI, 0.95–1.38). When stratified by sex, the RR of every 100 g/d increase in red meat consumption was significant in men (RR, 1.43; 95% CI, 1.40–1.86) but not significant in women (RR, 1.06; 95% CI, 0.86–1.30). Individuals who consumed processed meat showed a 17% increase in the risk of pancreatic cancer (RR, 1.17; 95% CI, 1.01–1.34), especially in men (RR, 1.21; 95% CI, 1.01–1.45), for every increase in intake of 50 g/day. When the intake of saturated fat increased by 10 g/d, the risk of pancreatic cancer increased by 11% (RR, 1.11; 95% CI, 1.01–1.21) [42].

3.2.2. Sugars, sugar-sweetened foods, and soft drinks

High intake of sugars, sugar-sweetened foods, or soft drinks also increase the risk of pancreatic cancer [43–45]. A pooled analysis of 14 cohort studies showed that the risk of pancreatic cancer was increased by 7% (RR, 1.07; 95% CI, 1.02–1.03) for the consumption of carbonated beverages with a daily sugar content of >175 g/day [45]. However, a recent study showed no significant correlation between sweet-beverage consumption and the risk of pancreatic cancer [46]. The association between coffee consumption and pancreatic cancer also remains controversial. A recent study showed that coffee intake was a protective factor for pancreatic cancer (moderate consumer >1 to <4 cups/day): hazard ratio (HR), 0.79; 95% CI, 0.64–0.99; heavy consumer (>4 cups/day): HR, 0.74; 95% CI, 0.57–0.95 [47]. However, the results of several published meta-analyses are contradictory and have not yielded a consistent conclusion [48–50]. Many experts believe that chronic pancreatitis might be a confounding factor in these studies; therefore, a high-quality meta-analysis is needed to clarify the relationship between coffee consumption and pancreatic cancer [51,52].

3.2.3. Other dietary factors

A recent large prospective cohort in Japan reported that soy products might increase the risk of pancreatic cancer, with an RR of 1.48 (95% CI, 1.15–1.92) [53], a finding that requires further confirmation. Vegetables, fruits, nuts, and whole-grain intake could reduce the risk of pancreatic cancer [54–57]. Meanwhile, the protective effect of vitamin D on pancreatic cancer remains controversial [58,59].

3.2.4. Alcohol

Population studies showed that alcohol consumption increased the risk of pancreatic cancer. The higher the alcohol consumption, the higher the risk of pancreatic cancer [60]. A meta-analysis showed that the risk of pancreatic cancer increased by 15% (RR, 1.15; 95% CI, 1.06–1.25) in heavy drinkers (average alcohol consumption \geq 24 g/d), while the risk in heavy liquor drinkers was quite high (RR, 1.43; 95% CI, 1.17–1.74) [61]. In addition, alcohol consumption was negatively correlated with survival time after pancreatic cancer diagnosis. The results of cohort studies showed that every 10 g/day increase in alcohol intake increased the risk of death due to pancreatic cancer by 9% (HR, 1.09; 95% CI, 1.00–1.19) [62]. Animal experiments also confirmed that moderate drinking promoted pancreatic cancer progression in mice [63].

3.2.5. Smoking

Smoking is also closely associated with pancreatic cancer occurrence and development. The risks of pancreatic cancer in current (RR, 1.8; 95% CI, 1.7–1.9) and former (RR, 1.2; 95% CI, 1.1–1.2) smokers was significantly higher than that in non-smokers [64]. The higher the amount of smoking, the higher the risk of pancreatic cancer. Smoking more than 30 cigarettes a day results in an increase in RR to 2.2 (95% CI, 1.9–2.4); correspondingly, quitting smoking helped to prevent pancreatic cancer (RR, 0.6; 95% CI, 0.5–0.7) [64]. Smoking can also significantly increase the risk of death due to pancreatic cancer. The results of a large cohort study showed that current smokers had a 37% higher risk of pancreatic cancer than did non-smokers (RR, 1.37; 95% CI, 1.11–1.69), with even higher risks observed in heavy smokers (RR, 1.76; 95% CI, 1.23–2.51) [65].

3.2.6. Physical activity

Appropriate physical activity may have a protective effect against pancreatic cancer. A meta-analysis of 14 studies showed that people who performed at least 150 min of physical activity at a certain intensity every week have 15% reduced risk of pancreatic cancer (RR, 0.85; 95% CI, 0.78–0.93) [66]. Another meta-analysis including 30 different studies reported that physical activity risk estimates appeared to be more pronounced for consistent physical activity over time (RR, 0.86; 95% CI, 0.76–0.97) than for recent-past (RR, 0.95; 95% CI, 0.90–1.01) or distant-past (RR 0.95; 95% CI, 0.79–1.15) physical activity [67].

3.2.7. Overweight and obesity

Overweight and obesity, as a risk factor for multiple metabolic disorders, can also increase the risk of pancreatic cancer, the mechanisms of which may be associated with inflammation, microbiota, hormones, and high-fat diets [68]. A meta-analysis showed that obesity (body mass index [BMI] \geq 30 kg/m²) may increase the risk of pancreatic cancer by 34–36% (male RR, 1.36; 95% CI, 1.07–1.73; female RR, 1.34; 95% CI, 1.22–1.46) [69]; however, the prognosis of patients with pancreatic cancer was not affected [70]. At the same time, overweight and obesity in adolescence may have a long-term effect on the risk of pancreatic cancer. A study of 1.79 million Israeli adolescents followed for more

than 20 years showed that compared to normal weight (5th to <85th percentiles), obesity (\geq 95th percentile) was associated with an increased risk of pancreatic cancer among both men (HR, 3.67; 95% CI, 2.52–5.34) and women (HR, 4.07; 95% CI, 1.78–9.29), with an estimated population-attributable fraction due to overweight and obesity of 10.9% (95% CI, 6.1–15.6%) [71]. In addition, the results of a randomized controlled trial (RCT) also showed that a low-fat diet intervention reduced the risk of pancreatic cancer in overweight women (BMI \geq 25 kg/m²) (HR, 0.71; 95% CI, 0.53–0.96) [72].

3.2.8. Aspirin use

Aspirin is a nonsteroidal anti-inflammatory drug used primarily for the prevention and treatment of cardiovascular diseases; however, its preventive effect in pancreatic cancer also has been reported [73,74]. A study including 761 patients with pancreatic cancer and 794 healthy individuals showed that ever-regular use of aspirin was associated with a lower risk of pancreatic cancer (OR, 0.54; 95% CI, 0.40–0.73) and that the risk decreased by 8% with each cumulative year of use (OR_{trend}, 0.92; 95% CI, 0.87–0.97) [73]. A meta-analysis published in 2020 that included 15 original studies showed that aspirin use reduced the risk of pancreatic cancer, with a pooled RR of 0.78 (95% CI, 0.68–0.89) [74]. However, recent cohort studies reported non-significant results; for example, Natalia Khalaf et al. [75] and Risch et al. [76] both reported no significant association between aspirin use and the risk of pancreatic cancer.

3.2.9. Exposure to trace elements

Trace elements, such as iron and cadmium, are reportedly associated with an increased risk of pancreatic cancer. A prospective study showed an association between heme–iron and increased pancreatic cancer risk in female smokers, with an HR (per 1 mg/day increase) of 1.38 (95% CI, 1.10–1.74), which increased significantly to 2.5-fold (95% CI, 1.22–5.28) after calibration [77]. Urinary cadmium concentrations were also significantly associated with an increased risk of pancreatic cancer (2nd quartile OR, 3.34; 3rd, 5.58; 4th, 7.70; test for trend p < 0.0001) [78]. In addition, exposure to trace elements from the environment, such as arsenic (HR, 2.46; 95% CI, 1.09–5.58) [79] and lead (OR, 6.26; 95% CI, 2.71–14.47) may also increase pancreatic cancer risk [80]. Conversely, selenium and nickel concentrations were negatively associated with the risk of pancreatic cancer [80]; however, this association requires further verification.

3.3. Disease status

The occurrence and development of pancreatic cancer are related to chronic pancreatitis, pancreatic cystic lesions, type II diabetes, infection, and other diseases.

3.3.1. Chronic pancreatitis

Chronic pancreatitis is a pathological change in pancreatic tissue and is considered an important risk factor for pancreatic cancer [81]. A recent meta-analysis showed a 16-fold increased lifetime risk of pancreatic cancer in patients with chronic pancreatitis (RR, 16.16; 95% CI, 12.59–20.73) [82]. The main manifestations of chronic pancreatitis are the destruction of pancreatic acinar cells and pathological fibrosis. A series of inflammatory processes, supplemented by somatic cells and genetic mutations, increase the risk of pancreatic cancer. Patients with genetically determined idiopathic chronic pancreatitis have a higher risk of pancreatic cancer, while the risk of alcohol-related chronic pancreatitis may be much lower [83]. However, although there is a strong association between chronic pancreatitis and pancreatic cancer, less than 5% of patients develop pancreatic cancer [84].

3.3.2. Pancreatic cystic lesions

Pancreatic cystic lesions include pancreatic pseudocysts (PPC) and pancreatic cystic neoplasms (PCN), the latter of which are a heterogeneous group of pancreatic cysts, such as intraductal papillary mucinous neoplasms (IPMN), mucinous cystic neoplasms (MCN), serous cystic neoplasms (SCN), and other rare cystic lesions, among which IPMN is the most common. Among these, IPMN and MCN are regarded as precursors to pancreatic cancer [85]. The risk of advanced neoplasia in IPMN is increased predominantly by main duct involvement, with a mean frequency of 62% in resected specimens. Individuals with IPMN also have an increased risk (1–8%) of developing conventional pancreatic ductal adenocarcinoma (PDAC) elsewhere in the pancreas [86]. A 10–39% increased risk of advanced neoplasia has been reported in patients with resected MCN [87–92]. Up to 15% and 10% of patients with resected solid pseudopapillary neoplasms (SPN) and cystic neuroendocrine tumors (cNET) develop invasive cancer, respectively [93,94].

3.3.3. Type II diabetes

Type II diabetes, characterized by insulin resistance, insufficient secretion, and abnormal glucose metabolism, is also closely associated with pancreatic cancer. A meta-analysis showed a 2-fold increase in the risk of pancreatic cancer in patients with type II diabetes (RR, 1.94; 95% CI, 1.66–2.27) and that the relative risk of pancreatic cancer was negatively correlated with the duration of diabetes [95]. Moreover, the risk of pancreatic cancer increased more significantly when newly diagnosed diabetes patients experienced recent weight loss, with an HR as high as 6.75 (95% CI, 4.55–10.00) [3]. A meta-analysis also showed that every 0.56 mmol/L (10 mg/dL) increase in fasting blood glucose level was associated with a 14% increased risk of pancreatic cancer (95% CI, 1.06–1.22) [9]. However, damage to pancreatic tissue affects insulin secretion function, which in turn may cause diabetes, thus demonstrating the complex relationship between pancreatic cancer and diabetes.

3.3.4. Infection

Some microbial infections, such as hepatitis B virus and *Helicobacter pylori*, mainly those of the digestive tract, have been associated with an increased risk of pancreatic cancer [96–98]. The estimated adjusted ORs and 95% CIs for hepatitis B virus and hepatitis C virus were as follows: anti-HCV-positive, 0.9 (95% CI, 0.3–2.8); anti-HBc-positive, 2.5 (95% CI, 1.5–4.2); anti-HBc-positive/anti-HBs-positive, 2.3 (95% CI, 1.2–4.2); and anti-HBc-positive/anti-HBs-negative, 4 (95% CI, 1.4–11.1) [99]. Regarding *H. pylori* infection, compared to seronegative subjects, those positive for *H. pylori* or cytotoxin-associated gene-A-positive strains showed a significantly increased risk of pancreatic cancer (OR, 1.87; 95% CI, 1.05–3.34; OR, 2.01; 95% CI, 1.09–3.70, respectively) [100].

4. Pancreatic cancer risk prediction models

The risk prediction models for pancreatic cancer can help to identify high-risk individuals for further intervention. Table 2 summarizes relevant studies on pancreatic cancer risk models.

The PancPRO [101] was the first risk prediction model for FPC and provided mutation carrier probability and absolute risk for a specified age interval. It included 6134 individuals from 961 families to establish a model based on the Mendelian risk prediction approach. This model has a relatively satisfactory performance, with an area under the curve (AUC) of 0.75. The PancPRO scored families based on pedigree data and assigned a quantitative risk score to any family member depending on the age at diagnosis (or death) of the affected relatives.

Another two studies also developed risk prediction models for the general population [102,103]; however, the performances of these models were not satisfactory, with both AUCs <0.7. In recent years, several teams have established risk prediction models in patients with new-onset diabetes (NOD) [8,103–105], most of which performed well. Notably, the risk prediction model of pancreatic cancer for the NOD patients conducted by Ayush Sharma et al. [8] had the highest AUC (0.87). The model was also convenient to use because it included only three parameters: change in weight, change in blood glucose level, and

Table 2

Summary of studies on pancreatic cancer risk models.

Study Source	Study Population	Sample Size	Study Design	Predictive factors	Prediction approach	AUC
Aileen Baecker et al. [103], 2019	General population and NOD	88938	Case-control study	Race, presence of at least 1 claim for acute pancreatitis, chronic pancreatitis, diabetes mellitus, dyspepsia, gallbladder disease, any abdominal pain, weight loss, jaundice, influenza vaccination, depression, chest pain	Logistic regression model	0.683(general population); 0.735(NOD)
Ayush Sharma et al. [8], 2018	NOD	1561	Retrospective cohort study	Change in weight, change in blood glucose, age at DM onset	Logistic regression model	0.87
Xin Dong et al. [104], 2018	NOD	413	Matched case- control study	BMI, age of DM onset, HBV infection, TBIL, ALB, ALT, BUN, Cr, TC, TG, HDL, LDL, APO-A1, APO-B, WBC count, HB, PLT count, PT	Logistic regression model	0.82
Ben Boursi et al. [105], 2017	NOD	109385	Retrospective cohort study	Age, BMI, change in BMI, smoking, insulin, oral hypoglycemics (not metformin), metformin, PPIs, HbA1c, Hb, cholesterol, creatinine, alkaline phosphatase	Logistic regression model	0.82
Alison P. Klein et al. [102], 2013	General population	7018	Case-control study	Current smoking, heavy alcohol use, obesity, diabetes >3 years, family history of pancreatic cancer, O/ABO genotype, rs3790844 (chr1q32.1), rs401681 (5p15.33), rs9543325 (13q22.1)	Logistic regression model	0.58(only non-genetic factors); 0.57(only genetic factors); 0.61 (both non-genetic and genetic factors)
Wenyi Wang et al. [101], 2007	FPC	6134	Prospective cohort	Pedigree data, age of family members combined with knowledge of the genetic transmission of pancreatic cancer.	Mendelian Risk Prediction Model	0.75

‡:NOD, New-Onset Diabetes; DM, Diabetes Melitums; FPC, Familial Pancreatic Cancer; BMI, Body Mass Index; HBV, Hepatitis B Virus; TBIL, Total Bilirubin; ALB, Albumin; ALT, Alanine Aminotransferase; BUN, Burea Nitrogen; Cr, Creatinine; TC, Total Cholesterol; TG, Total Glycerin Three Greases; HDL, High-Density Lipoprotein ; LDL, Low-Density Lipoprotein; APO-A1, Apolipoprotein-A1; APO-B, Apolipoprotein-B; WBC, White Blood Cell; HB, Hemoglobin; PLT, Platelet; PT, Prothrombin Time; HbA1c, Glycated Hemoglobin.

age at diabetes onset. However, research regarding risk prediction models for pancreatic cancer remain unsatisfactory both in quantity and quality; thus, additional studies are required.

5. Early detection and screening

5.1. Early detection modalities and biomarkers

Due to the lack of typical clinical manifestations and effective diagnostic methods, most patients with pancreatic cancer are in an advanced stage when diagnosed, with a low resection probability and poor treatment effect. Therefore, appropriate early diagnosis and screening strategies are particularly important for the early identification of pancreatic cancer patients to increase surgical opportunities and provide earlier treatment. At present, the methods used for the early diagnosis of pancreatic cancer in the clinical setting mainly include computed tomography (CT), magnetic resonance imaging (MRI), endoscopic ultrasonography (EUS), endoscopic retrograde cholangiopancreatography (ERCP), and magnetic resonance cholangiopancreatography (MRCP) [106,107]; however, these methods are not sensitive enough to identify patients with early pancreatic cancer.

Biomarkers may play an important role in the early detection and screening of individuals at a high risk of pancreatic cancer. Many studies have explored potential biomarkers for the early-stage detection of pancreatic cancer, mainly including proteomic, metabolomic, genetic, or transcriptomic biomarkers [85]. Carbohydrate antigen 19-9 (CA19-9), carcinoembryonic antigen (CEA), and CA12-5 are the most commonly used biomarkers for pancreatic cancer in the clinical setting; however, their accuracy for the early diagnosis of pancreatic cancer is not satisfactory [4,108,109]. The sensitivity and specificity of CA19-9 in the diagnosis of symptomatic patients can reach 79-81%, and 82-90%, respectively [110]. Therefore, many teams are committed to exploring and developing new biomarkers for early diagnosis. Recently, liquid biopsy has shown promise as a research direction for screening biomarkers. Researchers have screened exosomes, miRNAs, proteins, lipid metabolites, and other early diagnostic biomarkers of pancreatic cancer from body fluids including blood, saliva, urine, and pancreatic juice,

including lymphatic vessel endothelial hyaluronic acid receptor 1 (LYVE1), regenerating family member 1 beta (REG1B), and trefoil factor 1 (TFF1) protein levels in urine combined with CA19-9 in serum [111], and mucin 1 (MUC1) and MUC2 in pancreatic juice [112], etc [6,113, 114]. Besides proteins, several RNA biomarkers also showed potential, including miR-1246 [115], miR-25 combined with CA19-9 [116], miR-1290 combined with CA19-9 [117], and ABHD11-AS1 combined with CA19-9 or CEA or CA125 [118]. In addition, CancerSEEK [119], which uses combined assays for genetic alterations and protein biomarkers, and neutrophil-to-lymphocyte ratio (NLR) [120] also showed relatively satisfactory performances. Detailed information of studies on these potential biomarkers is shown in Table 3.

5.2. Screening

Screening is an important method to detect patients with early pancreatic cancer. However, it is not recommended for the general population, mainly due to the low incidence of pancreatic cancer and the low screening benefit. In addition, the accuracy of available screening methods is not satisfactory and some may also have negative effects on human health, such as pain and anesthesia-related adverse reactions after EUS examination, acute pancreatitis, and even hospitalization after ERCP, as well as anxiety and psychological effects [107]. Regarding populations that are at a high of pancreatic cancer, many teams are still evaluating appropriate screening strategies. The International Federation of Pancreatic Screening (CAPS) published guidelines for pancreatic cancer screening in 2012 [121], recommending screening out for high-risk groups with a lifetime risk of pancreatic cancer of >5% or more than five times the RR. The definition of high-risk population mainly refers to the population with a strong family history or genetic predisposition, including familial pancreatic cancer (FPC), genetic syndrome (LS, PJS, and gene mutations (p16, BRCA2, and PALB2). EUS and MRI are the most accurate and non-invasive tools used for the early screening of pancreatic cancer. In recent years, the United States [122-126], the Netherlands [127], Denmark [128], Germany [129], Sweden [130], Canada [131], and other countries have conducted screening programs for pancreatic cancer in high-risk groups. They mainly enrolled

Table 3

Selected studies on potential biomarkers for early detection of pancreatic cancer, published between 2017 and 2021.

Study Source	Biomarkers panel	Sample size	Marker type	Sample	AUC
Debernardi et al. [111], 2020	LYVE1, REG1B, TFF1, and CA19-9	PC (n = 199); BHD (n = 208); HC (n = 183)	Protein	Urine (LYVE1, REG1B, TFF1); serum (CA19-9)	0.992 (PDAC stage I-II vs. HC); 0.919 (PDAC stage I-II vs. Benign)
Tanaka et al. [112], 2019	MUC1 and MUC2	193 studies, comprising 12297 patients	Protein	Pancreatic juice	0.85 (malignant vs. benign IPMN)
Lee et al. [113], 2018	CEMIP	PC (n = 324); HC (n = 49); PC I-II (n = 88)	Protein	Serum	0.94 (PDAC vs. benign disease and HC)
Kim et al. [114], 2017	THBS-2 and CA19-9	PC III-IV (n = 109); HC (n = 140); PC I (n = 16)	Protein	Plasma	0.97 (PDAC all stages vs. HC); 0.96 (PDAC I/II vs. HC)
Mellby et al. [6], 2018	29-protein biomarker panel	PC II $(n = 132)$; PC III $(n = 65)$; PC IV $(n = 230)$; HC $(n = 888)$	Protein	Serum	0.96 (PDAC stage I-II vs. HC)
Ishige et al. [115], 2020	MiR-1246	PC (n = 41); HC (n = 30)	miRNA	Serum, urine and saliva	0.87 (serum, PDAC stage 0-IV vs. HC); 0.90 (urine, PDAC stage 0-IV vs. HC); 0.48 (saliva, PDAC stage 0-IV vs. HC)
Yu et al. [116], 2020	MiR-25 and CA19-9	PC (n = 80); HC (n = 90)	miRNA	Plasma	0.985 (PDAC stage I-IV vs. HC)
Tavano et al. [117], 2018	MiR-1290 and CA19-9	PC (n = 167); HC (n = 267)	miRNA	Plasma	0.956 (PDAC stage I-IV vs. HC)
Liu et al. [118], 2019	ABHD11-AS1 and CA19-9 or CEA or CA125	PC (n = 114); CP (n = 97); HC (n = 46)	lncRNA and protein	Plasma	0.982 (ABHD11-AS1+ CA19-9); 0.943 (ABHD11- AS1+ CEA); 0.914 (ABHD11-AS1+ CA125)
Cohen et al. [119], 2018	CancerSEEK	1005 cancer patients	DNA and proteins	Plasma	Sensitivity over 70% when specificity is over 99% (PDAC vs. HC)
Gemenetzis et al. [120], 2017	NLR	IPMN (n = 272)	Other	Serum	0.89 (non-invasive vs. invasive IPMN)

THBS2, Thrombospondin-2; PC, Pancreatic Cancer; HC, Healthy Controls; BHD, Benign Hepatobiliary Disease; CP, Chronic Pancreatitis; NLR, Neutrophil-to-Lymphocyte Ratio; CEMIP, Cell Migration-inducing hyaluronan Binding Protein.

individuals with a family history of pancreatic cancer and/or gene mutations and genetic syndromes. EUS and or MRI were the main examination methods, supplemented by CT, MRCP, and ERCP (details in Table 4).

For example, in 2006, Canto et al. performed a screening study [126] of 78 subjects with a family history of pancreatic cancer and PJS (the high-risk group) and 161 subjects in the control group. CT or ERCP combined with EUS was used for the examination. A total of 17 neoplastic-type lesions (21.8%) were detected in the high-risk group, compared to only one small cystic lesion (0.6%) in the control group. Finally, seven patients underwent surgical resection. Another screening study of 411 high-risk subjects with pancreatic cancer with a family history of pancreatic cancer and CDKN2A/p16, BRCA1/2, or PALB2 mutations published by Vasen et al., in 2016 [129] reported a total of 14 cases of pancreatic cancer, 2 cases of suspected pancreatic cancer, and 140 cases of pancreatic cystic lesions detected by MRI/MRCP combined with EUS, with a diagnosed yield of pancreatic cancer of 7.3%. Among these, 12 cases of pancreatic cancer were detected in the CDKN2A/p16 mutation group, with a resection rate of 75% and a 5-year survival rate of 24%, which was significantly higher than that of sporadic pancreatic cancer and symptomatic pancreatic cancer patients with the CDKN2A/p16 gene [132]. Precursor lesions were much more frequent in patients with FPC (52.4%) than in CDKN2A/p16-Leiden mutation carriers, while the yield of pancreatic cancer was very low (0.9%) [129].

Despite the focus on the high-risk population with FPC and genetic predisposition, some researchers have also proposed screening for populations at a high risk of sporadic pancreatic cancer based on risk enrichment due to new-onset diabetes, weight change, and other risk factors [8,133]. However, related research and evidence are limited. More studies are needed to identify populations that are at a high risk of pancreatic cancer and further support screening for pancreatic cancer.

6. Prognosis

The 5-year survival rate has improved slightly worldwide with the continuous development of treatment technology for pancreatic cancer. According to the SEER database, the overall 5-year relative survival rate during 2011–2017 was 10.8% in the US [1], compared to 14.3% in Australia, 11.4% in Canada, 9.88% in Norway, 9.59% in Denmark, and

7.93% in the United Kingdom in 2014 [134], and 7.2% in China during 2012–2015 [135].

The prognosis of patients with pancreatic cancer is not only related to tumor characteristics such as size, invasion site, molecular typing, TNM stage but is also affected by patient status and treatment. In the US, approximately half of the patients had distant-stage disease, with a 5year relative survival rate of only 2.9%, while 11% of patients had localized-stage tumors and a 5-year relative survival rate of 39.4% [1]. Bailey et al. [136] defined four pancreatic cancer subtypes, viz., squamous, pancreatic progenitor, immunogenic, and aberrantly differentiated endocrine exocrine, while Collisson et al., in 2011 [137] defined three subtypes, viz., classical, quasi-mesenchymal (OM-PDA), and exocrine-like type. Among these, the squamous, QM-PDA, and basal-like subtypes were associated with poor prognosis [136,137]. Several biomarkers predicting patient prognosis have been reported in recent years. For example, the presence of mutated KRAS [138], ATM and TP53 [139], and protein arginine methyltransferase 1 (PRMT1) [140] was correlated with worse prognosis. Wang et al., in 2019 [141] reported SCAMP1, HCP5, MAL2, and LINC00511 as key long non-coding RNAs (lncRNAs) predicting prognosis. Yokoyama et al., in 2020 [142] also reported that mucins (MUC) played crucial roles in carcinogenesis and tumor invasion in pancreatic cancers and also developed a machine-learning prognosis prediction model. However, high-quality clinical studies with large samples are still needed for further verification in this field.

7. Summary and future perspectives

As a malignant tumor with poor prognosis, improving the overall survival rate of pancreatic cancer is a major challenge. In recent years, the incidence of pancreatic cancer has been rising worldwide, especially in younger individuals. Elucidating the underlying mechanism of the tumorigenesis of pancreatic cancer, effective control of risk factors, and implementation of effective early screening and detection techniques may help to reduce disease burden. The risk factors for pancreatic cancer mainly include those related to individual characteristics, lifestyle behaviors, environment, and disease status. These provide a certain direction for the etiological prevention and screening of pancreatic cancer. However, additional study of the causal relationships is required.

Table 4

Results of selected prospective pancreatic cancer screening studies.

Study Source	Recruitment Period	Country	Eligible for inclusion	No. of Participants screened	Age, Mean (SD) [Range], y	Screening Methodology	Lesions detected	Management
Barnes et al. [122], 2018	2012–2017	USA	FPC; PJS; gene panels including APC, ATM, BMPR1A, BRCA1, BRCA2, CDKN2A, CDK4, EPCAM, FANCC, MLH1, MSH2, MSH6, NF1, PALB2, PALLD, PMS2, SMAD4, STK11, TP53	65	56 (14) [NR]	MRI; EUS after positive in MRI	28 patients with lesions by MRI; 9 of 28 were detected by EUS	-
Gangi et al. [123], 2018	2007–2017	USA	FPC; PJS; HP; FAMMM; <i>BRCA2</i> mutation	58	60 (NR) [NR]	EUS	15 Hyperechoic foci;8 Fat stranding; 5 Lobularity; 3 Hyperechoic main pancreatic duct; 2 Calcifications; 1 Pancreatic ductal dilation	Positive subjects get further FNA
Harinck et al. [127], 2016	2006–2013	Netherlands	FPC; PJS; mutation of <i>CDKN2A,</i> <i>BRCA1, BRCA2,</i> <i>p53</i>	139 at baseline;	51.1 (9.7) [20- 73]	EUS and MRI	Baseline:135 at the 2nd round; 2 solid lesions, 9 cysts ≥10 mm; 2nd round:12 solid lesions in 8 individuals were detected	Interval 3 or 6 months; Standard FU at 12 months; 12 lesions in 8 individuals were detected after 12 months (2nd round)
Joergensen et al. [128], 2016	2006–2014	Denmark	FPC; mutation of <i>PRSS1</i>	71	51.1 (NR) [26–72]	EUS	2 PDAC	Surgical resection (n = 2)
Vasen et al. [129], 2016	2002–2009	Germany	FPC; mutation of CDKN2A, p16, BRCA1/2, PALB2	CDKN2A:178 FPC: 214 BRCA1/2 or PALB2:19	CDKN2A:56 (NR) [37–75] FPC::48.2 (NR) [27–81] BRCA1/ 2 or PALB2:52.6 (NR) [25–70]	EUS and MRI/ MRCP	CDKN2A: PDAC (n = 13), cystic lesion (n = 26); FPC: suspected PDAC (n = 3), cystic lesion (n = 112); BRCA1/2 or PALB2: PDAC (n = 1), cystic lesion (n = 2)	Surgical resection (n = 31); surveillance
Del Chiaro et al. [130], 2015	2010–2013	Sweden	FPC; PJS; mutation of <i>BRCA2, BRCA1/</i> p16	40	49.9 (NR) [23–76]	MRI; EUS after positive in MRI	PDAC ($n = 2$); branch duct (BD) IPMN($n =$ 9); mixed-type IPMN ($n = 3$); main duct IPMN in ($n = 2$)	Surgical resection (n = 5); surveillance
Al-Sukhni et al. [131], 2012	2003–2011	Canada	FPC; PJS or HP; mutation of <i>p16,</i> <i>STK11, BRCA2,</i> <i>BRCA1</i>	262	54 (NR) [22–89]	MRI	3 PDAC; 15 BD IPMNs; 65 simple pancreatic cysts; 22 mildly dilated main pancreatic ducts	Surgery or surveillance
Canto et al. [124], 2012	2006–2009	USA	FPC; PJS; FBOC	216	56.1 (NR) [28–79]	EUS and CT and MRI/ MRCP	3 solid lesions; 84 cystic lesions; 5 isolated dilated main pancreatic ductother surveillance	5 Surgically- Treated
Ludwig et al. [125], 2011	2002–2009	USA	FPC; mutation of <i>BRCA</i>	109	54 (11.4) [33–86]	MRCP or CT for those unwilling to undergo MRCP	9 significant lesions	6 surgical resection; all under surveillance
Canto et al. [126], 2006	2001–2004	USA	FPC; PJS	High-risk: 78 Controls: 161	High-risk: 52 (NR) [32–77] Controls: 54 (NR) [30–80]	High-risk: EUS and CT; Controls: EUS and/or ERCP	High-risk: neoplastic- type lesions ($n = 17$); Controls: a small cyst ($n = 1$)	7 surgical resection; 9 surveillance; 1 FNA

§: FPC, Familial Pancreatic Cancer; FNA, Fine-Needle Aspiration; FAMMM, Familial-Atypical Multiple Mole Melanoma Syndrome; HP, Hereditary Pancreatitits; PJS, Peutz-Jeghers Syndrome; FBOC, Familial Breast Ovarian Cancer Syndrome; PCMS, Pancreatic Melanoma Cancer Syndrome.

Several prediction models for pancreatic cancer have been developed in populations with new-onset diabetes or a family history of pancreatic cancer and the general population; however, these require further validation. Despite recent screening efforts, the quantity and quality of related studies remains unsatisfactory, especially regarding the definitions of high-risk groups and screening modality design. Apart from the populations of individuals with familial genetic risk and those at a high risk of sporadic pancreatic cancer, risk factors such as new-onset diabetes, obesity, and others may provide a new direction for the screening of at-risk populations.

Increased attention and investment in the field of pancreatic cancer epidemiology are needed to provide more high-quality evidence for its prevention and control. High-quality cohorts of high-risk populations and a global sharing data platform of pancreatic cancer may be helpful in the development of this field.

CRediT authorship contribution statement

Jie Cai: Conceptualization, Writing – original draft. Hongda Chen: Conceptualization, Writing – review & editing. Ming Lu: Methodology. Yuhan Zhang: Visualization. Bin Lu: Visualization. Lei You: Conceptualization. Taiping Zhang: Supervision. Min Dai: Writing – review & editing, Supervision. Yupei Zhao: Supervision.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Abbreviations

GLOBOCA	AN Global Cancer Observatory					
ASR	age standardized rate					
AAPC	average annual percent change					
HDI	human development index					
DALYs	disability adjusted of life years					
SEER	Surveillance, Epidemiology, and End Results					
CI	confidence interval					
OR	odds ratio					
Cag	acytotoxin-associated gene A					
RR	relative risk					
HPC	hereditary pancreatic cancer					
PJS	Peutz-Jeghers syndrome					
HP	hereditary pancreatitis					
FAMMM	familial atypical mole and multiple melanoma					
HBOC	hereditary breast and ovarian cancer syndrome					
LS	Lynch syndrome					
FAP	familial adenomatous polyposis					
BRCA2	breast cancer gene 2					
PALB2	partner and localizer of BRCA2					
ATM	ataxia-telangiectasia-mutated					
APC	adenomatous polyposis coli					
MLH1	MutL homolog 1					
MSH2	MutS homolog 2					
PMS2	PMS1 homolog 2					
PRSS1	serine protease 1					
STK11	serine/threonine kinase 11					
CTRC	chymotrypsin C					
LFS	Li-Fraumeni syndrome					
AT	ataxia telangiectasia					
CF	Cystic fibrosis					
HR	hazard ratio					
BMI	body mass index					
RCT	randomized controlled trial					
PPC	pancreatic pseudocyst					
PCN	pancreatic cystic neoplasms					
IPMN	intraductal papillary mucinous neoplasms					
MCN	mucinous cystic neoplasms					
SCN	serous cystic neoplasms					
PDAC	pancreatic ductal adenocarcinoma					
SPN	solid pseudopapillary neoplasms					
cNET	cystic neuroendocrine tumors					
AUC	area under curve					
NOD	new-onset diabetes					
CT	computed tomography					
MRI	magnetic resonance imaging					
EUS	endoscopic ultrasonography					
ERCP	endoscopic retrograde cholangiopancreatography					
MRCP	magnetic resonance cholangiopancreatography					

- CA19-9 carbohydrate antigen 19-9
- CEA carcinoembryonic antigen
- CA-125 carbohydrate antigen-125
- LYVE1 lymphatic vessel endothelial hyaluronic acid receptor 1
- REG1B regenerating family member 1 beta
- TFF1 trefoil factor 1
- MUC1 mucin 1
- CEMIP cell migration-inducing hyaluronan binding protein
- NLR Neutrophil-to lymphocyte ratio
- FPC familial pancreatic cancer
- QM-PDA Quasi-mesenchymal
- PRMT1 protein arginine methyltransferase 1
- lncRNAs long non-coding RNAs
- MUC Mucins
- DM diabetes mellitus

References

- S.S.F. Sheets, Pancreas cancer. https://seer.cancer.gov/statfacts/html/pancreas. html. Accessed 2/18/2021.
- [2] H.L. O'Gradyand, K.C. Conlon, Pancreatic neuroendocrine tumours, Eur. J. Surg. Oncol. 3 (34) (2008) 324–332.
- [3] C. Yuan, A. Babic, N. Khalaf, et al., Diabetes, weight change, and pancreatic cancer risk, JAMA Oncol. 10 (6) (2020), e202948.
- [4] J.F. Fahrmann, C.M. Schmidt, X. Mao, et al., Lead-time trajectory of CA19-9 as an anchor marker for pancreatic cancer early detection, Gastroenterology 4 (160) (2021) 1373–1383, e1376.
- [5] F. Kastrinos, B. Mukherjee, N. Tayob, et al., Risk of pancreatic cancer in families with Lynch syndrome, J. Am. Med. Assoc. 16 (302) (2009) 1790–1795.
- [6] L.D. Mellby, A.P. Nyberg, J.S. Johansen, et al., Serum biomarker signature-based liquid biopsy for diagnosis of early-stage pancreatic cancer, J. Clin. Oncol. 28 (36) (2018) 2887–2894.
- [7] J. Huang, V. Lok, C.H. Ngai, et al., Worldwide burden of, risk factors for, and trends in pancreatic cancer, Gastroenterology 3 (160) (2021) 744–754.
- [8] A. Sharma, H. Kandlakunta, S.J.S. Nagpal, et al., Model to determine risk of pancreatic cancer in patients with new-onset diabetes, Gastroenterology 3 (155) (2018) 730–739, e733.
- [9] W.C. Liao, Y.K. Tu, M.S. Wu, et al., Blood glucose concentration and risk of pancreatic cancer: systematic review and dose-response meta-analysis, BMJ (350) (2015) g7371.
- [10] P. Moller, T.T. Seppala, I. Bernstein, et al., Cancer risk and survival in path_MMR carriers by gene and gender up to 75 years of age: a report from the Prospective Lynch Syndrome Database, Gut 7 (67) (2018) 1306–1316.
- [11] Globocan, Cancer today. https://gco.iarc.fr/today/home. (Accessed 10 January 2021). Accessed.
- [12] R.L. Siegel, K.D. MillerandA. Jemal, Cancer statistics, 2020, CA A Cancer J. Clin. 1 (70) (2020) 7–30.
- [13] L. Rahib, B.D. Smith, R. Aizenberg, et al., Projecting cancer incidence and deaths to 2030: the unexpected burden of thyroid, liver, and pancreas cancers in the United States, Canc. Res. 11 (74) (2014) 2913–2921.
- [14] P. Maisonneuve, Epidemiology and burden of pancreatic cancer, Presse Med. 3 Pt 2 (48) (2019) e113–e123.
- [15] S.W. Zhang, R.S. Zheng, H.M. Zeng, et al., Cancer incidence and mortality in China, 2015, J. Nat. Canc. Cent. 1 (1) (2021) 2–11.
- [16] G.P.C. Collaborators, The global, regional, and national burden of pancreatic cancer and its attributable risk factors in 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017, Lancet Gastroenterol. Hepatol. 12 (4) (2019) 934–947.
- [17] X.H. Xu, X.Y. Zeng, L.J. Wang, et al., The disease burden of pancreatic cancer in China in 1990 and 2017, Zhonghua Liuxingbingxue Zazhi 9 (40) (2019) 1084–1088.
- [18] S. Midha, S. ChawlaandP, K. Garg, Modifiable and non-modifiable risk factors for pancreatic cancer: a review, Canc. Lett. 1 (381) (2016) 269–277.
- [19] Y. Matsuda, Age-related morphological changes in the pancreas and their association with pancreatic carcinogenesis, Pathol. Int. 8 (69) (2019) 450–462.
- [20] M.M. Wahi, N. Shah, C.E. Schrock, et al., Reproductive factors and risk of pancreatic cancer in women: a review of the literature, Ann. Epidemiol. 2 (19) (2009) 103–111.
- [21] D.T. Silverman, R.N. Hoover, L.M. Brown, et al., Why do Black Americans have a higher risk of pancreatic cancer than White Americans? Epidemiology 1 (14) (2003) 45–54.
- [22] D.S. Longnecker, M.R. Karagas, T.D. Tosteson, et al., Racial differences in pancreatic cancer: comparison of survival and histologic types of pancreatic carcinoma in Asians, blacks, and whites in the United States, Pancreas 4 (21) (2000) 338–343.
- [23] B.M. Wolpin, P. Kraft, M. Gross, et al., Pancreatic cancer risk and ABO blood group alleles: results from the pancreatic cancer cohort consortium, Canc. Res. 3 (70) (2010) 1015–1023.
- [24] L. Amundadottir, P. Kraft, R.Z. Stolzenberg-Solomon, et al., Genome-wide association study identifies variants in the ABO locus associated with susceptibility to pancreatic cancer, Nat. Genet. 9 (41) (2009) 986–990.

- [25] G.M. Petersen, L. Amundadottir, C.S. Fuchs, et al., A genome-wide association study identifies pancreatic cancer susceptibility loci on chromosomes 13q22.1, 1q32.1 and 5p15.33, Nat. Genet. 3 (42) (2010) 224–228.
- [26] H.A. Risch, L. Lu, J. Wang, et al., ABO blood group and risk of pancreatic cancer: a study in Shanghai and meta-analysis, Am. J. Epidemiol. 12 (177) (2013) 1326–1337.
- [27] T.P. Yeo, Demographics, epidemiology, and inheritance of pancreatic ductal adenocarcinoma, Semin. Oncol. 1 (42) (2015) 8–18.
- [28] J. Permuth-WeyandK, M. Egan, Family history is a significant risk factor for pancreatic cancer: results from a systematic review and meta-analysis, Fam. Cancer 2 (8) (2009) 109–117.
- [29] J. Llach, S. CarballalandL. Moreira, Familial pancreatic cancer: current perspectives, Canc. Manag. Res. (12) (2020) 743–758.
- [30] M.G. van Lier, A. Wagner, E.M. Mathus-Vliegen, et al., High cancer risk in Peutz-Jeghers syndrome: a systematic review and surveillance recommendations, Am. J. Gastroenterol. 6 (105) (2010) 1258–1264.
- [31] C.A. Shelton, C. Umapathy, K. Stello, et al., Hereditary pancreatitis in the United States: survival and rates of pancreatic cancer, Am. J. Gastroenterol. 9 (113) (2018) 1376.
- [32] V. Rebours, M.C. Boutron-Ruault, M. Schnee, et al., Risk of pancreatic adenocarcinoma in patients with hereditary pancreatitis: a national exhaustive series, Am. J. Gastroenterol. 1 (103) (2008) 111–119.
- [33] R.H. Hruban, M.I. Canto, M. Goggins, et al., Update on familial pancreatic cancer, Adv. Surg. (44) (2010) 293–311.
- [34] H.F. Vasen, N.A. Gruis, R.R. Frants, et al., Risk of developing pancreatic cancer in families with familial atypical multiple mole melanoma associated with a specific 19 deletion of p16 (p16-Leiden), Int. J. Canc. 6 (87) (2000) 809–811.
- [35] C. Hu, S.N. Hart, E.C. Polley, et al., Association between inherited germline mutations in cancer predisposition genes and risk of pancreatic cancer, J. Am. Med. Assoc. 23 (319) (2018) 2401–2409.
- [36] J.M. J. Mersch, M. Park, D. Nebgen, S.K. Peterson, C. Singletary, B.K. Arun, J. K. Litton, Cancers associated with BRCA1 and BRCA2 mutations other than breast and ovarian Cancer 14 (121) (2015) 2474–2475.
- [37] F.M. Giardiello, G.J. Offerhaus, D.H. Lee, et al., Increased risk of thyroid and pancreatic carcinoma in familial adenomatous polyposis, Gut 10 (34) (1993) 1394–1396.
- [38] N.J. Roberts, A.L. Norris, G.M. Petersen, et al., Whole genome sequencing defines the genetic heterogeneity of familial pancreatic cancer, Canc. Discov. 2 (6) (2016) 166–175.
- [39] S. Naudin, V. Viallon, D. Hashim, et al., Healthy lifestyle and the risk of pancreatic cancer in the EPIC study, Eur. J. Epidemiol. 10 (35) (2020) 975–986.
- [40] M. Song, W.S. GarrettandA, T. Chan, Nutrients, foods, and colorectal cancer prevention, Gastroenterology 6 (148) (2015) 1244–1260, e1216.
- [41] J.L. Petrick, N. Castro-Webb, H. Gerlovin, et al., A prospective analysis of intake of red and processed meat in relation to pancreatic cancer among african American women, Canc. Epidemiol. Biomark. Prev. 9 (29) (2020) 1775–1783.
- [42] W.C.R.F. I, Continuous update project expert report 2018, Diet, Nutr. Phys. Activity Pancreat. Canc. (2018) 11–12.
- [43] S.C. Larsson, L. BergkvistandA. Wolk, Consumption of sugar and sugar-sweetened foods and the risk of pancreatic cancer in a prospective study, Am. J. Clin. Nutr. 5 (84) (2006) 1171–1176.
- [44] N.T. Mueller, A. Odegaard, K. Anderson, et al., Soft drink and juice consumption and risk of pancreatic cancer: the Singapore Chinese Health Study, Canc. Epidemiol. Biomark. Prev. 2 (19) (2010) 447–455.
- [45] J.M. Genkinger, R. Li, D. Spiegelman, et al., Coffee, tea, and sugar-sweetened carbonated soft drink intake and pancreatic cancer risk: a pooled analysis of 14 cohort studies, Canc. Epidemiol. Biomark. Prev. 2 (21) (2012) 305–318.
- [46] E.M. Navarrete-Muñoz, P.A. Wark, D. Romaguera, et al., Sweet-beverage consumption and risk of pancreatic cancer in the European prospective investigation into cancer and nutrition (EPIC), Am. J. Clin. Nutr. 3 (104) (2016) 760–768.
- [47] M. Lukic, L.M. Nilsson, G. Skeie, et al., Coffee consumption and risk of rare cancers in Scandinavian countries, Eur. J. Epidemiol. 3 (33) (2018) 287–302.
- [48] K. Nie, Z. Xing, W. Huang, et al., Coffee intake and risk of pancreatic cancer: an updated meta-analysis of prospective studies, Minerva Med. 4 (107) (2016) 270–278.
- [49] F. Turati, C. Galeone, V. Edefonti, et al., A meta-analysis of coffee consumption and pancreatic cancer, Ann. Oncol. 2 (23) (2012) 311–318.
- [50] T.D. Li, H.W. Yang, P. Wang, et al., Coffee consumption and risk of pancreatic cancer: a systematic review and dose-response meta-analysis, Int. J. Food Sci. Nutr. 5 (70) (2019) 519–529.
- [51] L. MarkoandB. Tonje, Evidence on coffee consumption and pancreatic cancer: not great, not terrible, Eur. J. Epidemiol. 10 (35) (2020) 989–990.
- [52] T. Kawada, Coffee consumption and pancreatic cancer, Eur. J. Epidemiol. 10 (35) (2020) 987–988.
- [53] Y. Yamagiwa, N. Sawada, T. Shimazu, et al., Soy food intake and pancreatic cancer risk: the Japan public health center-based prospective study, Canc. Epidemiol. Biomark. Prev. 6 (29) (2020) 1214–1221.
- [54] G.A. Gaesser, Whole grains, refined grains, and cancer risk: a systematic review of meta-analyses of observational studies, Nutrients 12 (12) (2020).
- [55] J.M. Bae, E.J. LeeandG. Guyatt, Citrus fruit intake and pancreatic cancer risk: a quantitative systematic review, Pancreas 2 (38) (2009) 168–174.
- [56] L. NieuwenhuisandP, A. van den Brandt, Total nut, tree nut, peanut, and peanut butter consumption and the risk of pancreatic cancer in The Netherlands cohort study, Canc. Epidemiol. Biomark. Prev. 3 (27) (2018) 274–284.

- [57] Y. Bao, F.B. Hu, E.L. Giovannucci, et al., Nut consumption and risk of pancreatic cancer in women, Br. J. Canc. 11 (109) (2013) 2911–2916.
- [58] M. Waterhouse, H.A. Risch, C. Bosetti, et al., Vitamin D and pancreatic cancer: a pooled analysis from the pancreatic cancer case-control consortium, Ann. Oncol. 8 (26) (2015) 1776–1783.
- [59] Y. Bao, K. Ng, B.M. Wolpin, et al., Predicted vitamin D status and pancreatic cancer risk in two prospective cohort studies, Br. J. Canc. 9 (102) (2010) 1422–1427.
- [60] S. Naudin, K. Li, T. Jaouen, et al., Lifetime and baseline alcohol intakes and risk of pancreatic cancer in the European Prospective Investigation into Cancer and Nutrition study, Int. J. Canc. 4 (143) (2018) 801–812.
- [61] Y.T. Wang, Y.W. Gou, W.W. Jin, et al., Association between alcohol intake and the risk of pancreatic cancer: a dose-response meta-analysis of cohort studies, BMC Canc. (16) (2016) 212.
- [62] H. Jayasekara, D.R. English, A.M. Hodge, et al., Lifetime alcohol intake and pancreatic cancer incidence and survival: findings from the Melbourne Collaborative Cohort Study, Cancer Causes Control 4 (30) (2019) 323–331.
- [63] K. Asahina, S. Balog, E. Hwang, et al., Moderate alcohol intake promotes pancreatic ductal adenocarcinoma development in mice expressing oncogenic Kras, Am. J. Physiol. Gastrointest. Liver Physiol. 2 (318) (2020) G265–g276.
- [64] A. Lugo, G. Peveri, C. Bosetti, et al., Strong excess risk of pancreatic cancer for low frequency and duration of cigarette smoking: a comprehensive review and meta-analysis, Eur. J. Canc. (104) (2018) 117–126.
- [65] C. Yuan, V. Morales-Oyarvide, A. Babic, et al., Cigarette smoking and pancreatic cancer survival, J. Clin. Oncol. 16 (35) (2017) 1822–1828.
- [66] F. Xie, Y. You, J. Huang, et al., Association between physical activity and digestive-system cancer: an updated systematic review and meta-analysis, J. Sport Health Sci. 1 (10) (2021) 4–13.
- [67] G. Behrens, C. Jochem, D. Schmid, et al., Physical activity and risk of pancreatic cancer: a systematic review and meta-analysis, Eur. J. Epidemiol. 4 (30) (2015) 279–298.
- [68] P. Cascetta, A. Cavaliere, G. Piro, et al., Pancreatic cancer and obesity: molecular mechanisms of cell transformation and chemoresistance, Int. J. Mol. Sci. 11 (19) (2018).
- [69] M. Dobbins, K. DecorbyandB, C. Choi, The association between obesity and cancer risk: a meta-analysis of observational studies from 1985 to 2011, ISRN Prev. Med. (2013) 680536.
- [70] P. Téoule, E. Rasbach, H. Oweira, et al., Obesity and pancreatic cancer: a matched-pair survival analysis, J. Clin. Med. 11 (9) (2020).
- [71] L. Zohar, Y. Rottenberg, G. Twig, et al., Adolescent overweight and obesity and the risk for pancreatic cancer among men and women: a nationwide study of 1.79 million Israeli adolescents, Cancer 1 (125) (2019) 118–126.
- [72] L. Jiao, L. Chen, D.L. White, et al., Low-fat dietary pattern and pancreatic cancer risk in the women's health initiative dietary modification randomized controlled trial, J. Nat. Canc. Inst. 1 (110) (2018) 49–56.
- [73] H.A. Risch, L. Lu, S.A. Streicher, et al., Aspirin use and reduced risk of pancreatic cancer, Canc. Epidemiol. Biomark. Prev. 1 (26) (2017) 68–74.
- [74] C. Bosetti, C. Santucci, S. Gallus, et al., Aspirin and the risk of colorectal and other digestive tract cancers: an updated meta-analysis through 2019, Ann. Oncol. 5 (31) (2020) 558–568.
- [75] N. Khalaf, C. Yuan, T. Hamada, et al., Regular use of aspirin or non-aspirin nonsteroidal anti-inflammatory drugs is not associated with risk of incident pancreatic cancer in two large cohort studies, Gastroenterology 5 (154) (2018) 1380–1390, e1385.
- [76] M.H. Kim, S.M. Park, Y.H. Yun, et al., Aspirin does not prevent pancreatic cancer in a large asian cohort, Canc. Epidemiol. Biomark. Prev. 4 (28) (2019) 826–828.
- [77] E. Molina-Montes, P.A. Wark, M.J. Sánchez, et al., Dietary intake of iron, hemeiron and magnesium and pancreatic cancer risk in the European prospective investigation into cancer and nutrition cohort, Int. J. Canc. 7 (131) (2012) E1134–E1147.
- [78] B.G. Luckett, L.J. Su, J.C. Rood, et al., Cadmium exposure and pancreatic cancer in south Louisiana, 2012, J. Environ. Publ. Health (2012) 180186.
- [79] E. García-Esquinas, M. Pollán, J.G. Umans, et al., Arsenic exposure and cancer mortality in a US-based prospective cohort: the strong heart study, Canc. Epidemiol. Biomark. Prev. 11 (22) (2013) 1944–1953.
- [80] A.F. Amaral, M. Porta, D.T. Silverman, et al., Pancreatic cancer risk and levels of trace elements, Gut 11 (61) (2012) 1583–1588.
- [81] D. Malka, P. Hammel, F. Maire, et al., Risk of pancreatic adenocarcinoma in chronic pancreatitis, Gut 6 (51) (2002) 849–852.
- [82] J. Kirkegård, F.V. Mortensen, D. Cronin-Fenton, Chronic pancreatitis and pancreatic cancer risk: a systematic review and meta-analysis, Am. J. Gastroenterol. 9 (112) (2017) 1366–1372.
- [83] S. Midha, V. Sreenivas, M. Kabra, et al., Genetically determined chronic pancreatitis but not alcoholic pancreatitis is a strong risk factor for pancreatic cancer, Pancreas 10 (45) (2016) 1478–1484.
- [84] D. YadavandA, B. Lowenfels, The epidemiology of pancreatitis and pancreatic cancer, Gastroenterology 6 (144) (2013) 1252–1261.
- [85] S.P. Pereira, L. Oldfield, A. Ney, et al., Early detection of pancreatic cancer, Lancet Gastroenterol. Hepatol. 7 (5) (2020) 698–710.
- [86] N.C.M. van Huijgevoort, M. Del Chiaro, C.L. Wolfgang, et al., Diagnosis and management of pancreatic cystic neoplasms: current evidence and guidelines, Nat. Rev. Gastroenterol. Hepatol. 11 (16) (2019) 676–689.
- [87] K.T. Jang, S.M. Park, O. Basturk, et al., Clinicopathologic characteristics of 29 invasive carcinomas arising in 178 pancreatic mucinous cystic neoplasms with ovarian-type stroma: implications for management and prognosis, Am. J. Surg. Pathol. 2 (39) (2015) 179–187.

- [88] B.K. Goh, Y.M. Tan, Y.F. Chung, et al., A review of mucinous cystic neoplasms of the pancreas defined by ovarian-type stroma: clinicopathological features of 344 patients, World J. Surg. 12 (30) (2006) 2236–2245.
- [89] J.W. Park, J.Y. Jang, M.J. Kang, et al., Mucinous cystic neoplasm of the pancreas: is surgical resection recommended for all surgically fit patients? Pancreatology 2 (14) (2014) 131–136.
- [90] R.P. Reddy, T.C. Smyrk, M. Zapiach, et al., Pancreatic mucinous cystic neoplasm defined by ovarian stroma: demographics, clinical features, and prevalence of cancer, Clin. Gastroenterol. Hepatol. 11 (2) (2004) 1026–1031.
- [91] M.G. Sarr, H.A. Carpenter, L.P. Prabhakar, et al., Clinical and pathologic correlation of 84 mucinous cystic neoplasms of the pancreas: can one reliably differentiate benign from malignant (or premalignant) neoplasms? Ann. Surg. 2 (231) (2000) 205–212.
- [92] G. Zamboni, A. Scarpa, G. Bogina, et al., Mucinous cystic tumors of the pancreas: clinicopathological features, prognosis, and relationship to other mucinous cystic tumors, Am. J. Surg. Pathol. 4 (23) (1999) 410–422.
- [93] S.E. Lee, J.Y. Jang, D.W. Hwang, et al., Clinical features and outcome of solid pseudopapillary neoplasm: differences between adults and children, Arch. Surg. 12 (143) (2008) 1218–1221.
- [94] Y.X. Koh, A.Y. Chok, H.L. Zheng, et al., A systematic review and meta-analysis of the clinicopathologic characteristics of cystic versus solid pancreatic neuroendocrine neoplasms, Surgery 1 (156) (2014) 83–96 e82.
- [95] Q. Ben, M. Xu, X. Ning, et al., Diabetes mellitus and risk of pancreatic cancer: a meta-analysis of cohort studies, Eur. J. Canc. 13 (47) (2011) 1928–1937.
- [96] C. Song, J. Lv, Y. Liu, et al., Associations between hepatitis B virus infection and risk of all cancer types, JAMA Netw. Open 6 (2) (2019), e195718.
- [97] H.B. El-Serag, E.A. Engels, O. Landgren, et al., Risk of hepatobiliary and pancreatic cancers after hepatitis C virus infection: a population-based study of U. S. veterans, Hepatology 1 (49) (2009) 116–123.
- [98] X.Z. Chen, R. Wang, H.N. Chen, et al., Cytotoxin-associated gene A-negative strains of Helicobacter pylori as a potential risk factor of pancreatic cancer: a meta-analysis based on nested case-control studies, Pancreas 8 (44) (2015) 1340–1344.
- [99] M.M. Hassan, D. Li, A.S. El-Deeb, et al., Association between hepatitis B virus and pancreatic cancer, J. Clin. Oncol. 28 (26) (2008) 4557–4562.
- [100] R.Z. Stolzenberg-Solomon, M.J. Blaser, P.J. Limburg, et al., Helicobacter pylori seropositivity as a risk factor for pancreatic cancer, J. Nat. Canc. Inst. 12 (93) (2001) 937–941.
- [101] W. Wang, S. Chen, K.A. Brune, et al., PancPRO: risk assessment for individuals with a family history of pancreatic cancer, J. Clin. Oncol. 11 (25) (2007) 1417–1422.
- [102] A.P. Klein, S. Lindström, J.B. Mendelsohn, et al., An absolute risk model to identify individuals at elevated risk for pancreatic cancer in the general population, PloS One 9 (8) (2013), e72311.
- [103] A. Baecker, S. Kim, H.A. Risch, et al., Do changes in health reveal the possibility of undiagnosed pancreatic cancer? Development of a risk-prediction model based on healthcare claims data, PloS One 6 (14) (2019), e0218580.
- [104] X. Dong, Y.B. Lou, Y.C. Mu, et al., Predictive factors for differentiating pancreatic cancer-associated diabetes mellitus from common type 2 diabetes mellitus for the early detection of pancreatic cancer, Digestion 4 (98) (2018) 209–216.
- [105] B. Boursi, B. Finkelman, B.J. Giantonio, et al., A clinical prediction model to assess risk for pancreatic cancer among patients with new-onset diabetes, Gastroenterology 4 (152) (2017) 840–850, e843.
- [106] J.D. Mizrahi, R. Surana, J.W. Valle, et al., Pancreatic cancer, Lancet 10242 (395) (2020) 2008–2020.
- [107] N.B. Henrikson, E.J. Aiello Bowles, P.R. Blasi, et al., Screening for pancreatic cancer: updated evidence report and systematic review for the US preventive services task force, J. Am. Med. Assoc. 5 (322) (2019) 445–454.
- [108] L. van Manen, J.V. Groen, H. Putter, et al., Elevated CEA and CA19-9 serum levels independently predict advanced pancreatic cancer at diagnosis, Biomarkers 2 (25) (2020) 186–193.
- [109] Q. Meng, S. Shi, C. Liang, et al., Diagnostic accuracy of a CA125-based biomarker panel in patients with pancreatic cancer: a systematic review and meta-analysis, J. Canc. 17 (8) (2017) 3615–3622.
- [110] U.K. BallehaninnaandR, S. Chamberlain, The clinical utility of serum CA 19-9 in the diagnosis, prognosis and management of pancreatic adenocarcinoma: an evidence based appraisal, J. Gastrointest. Oncol. 2 (3) (2012) 105–119.
- [111] S. Debernardi, H. O'Brien, A.S. Algahmdi, et al., A combination of urinary biomarker panel and PancRISK score for earlier detection of pancreatic cancer: a case-control study, PLoS Med. 12 (17) (2020), e1003489.
- [112] M. Tanaka, M. Heckler, B. Liu, et al., Cytologic analysis of pancreatic juice increases specificity of detection of malignant IPMN-A systematic review, Clin. Gastroenterol. Hepatol. 11 (17) (2019) 2199–2211, e2121.
- [113] H.S. Lee, C.Y. Jang, S.A. Kim, et al., Combined use of CEMIP and CA 19-9 enhances diagnostic accuracy for pancreatic cancer, Sci. Rep. 1 (8) (2018) 3383.
- [114] J. Kim, W.R. Bamlet, A.L. Oberg, et al., Detection of early pancreatic ductal adenocarcinoma with thrombospondin-2 and CA19-9 blood markers, Sci. Transl. Med. 398 (9) (2017).

- [115] F. Ishige, I. Hoshino, Y. Iwatate, et al., MIR1246 in body fluids as a biomarker for pancreatic cancer, Sci. Rep. 1 (10) (2020) 8723.
- [116] Y. Yu, Y. Tong, A. Zhong, et al., Identification of Serum microRNA-25 as a novel biomarker for pancreatic cancer, Medicine (Baltim.) 52 (99) (2020), e23863.
- [117] F. Tavano, D. Gioffreda, M.R. Valvano, et al., Droplet digital PCR quantification of miR-1290 as a circulating biomarker for pancreatic cancer, Sci. Rep. 1 (8) (2018) 16389.
- [118] Y. Liu, W. Feng, W. Liu, et al., Circulating IncRNA ABHD11-AS1 serves as a biomarker for early pancreatic cancer diagnosis, J. Canc. 16 (10) (2019) 3746–3756.
- [119] J.D. Cohen, L. Li, Y. Wang, et al., Detection and localization of surgically resectable cancers with a multi-analyte blood test, Science 6378 (359) (2018) 926–930.
- [120] G. Gemenetzis, F. Bagante, J.F. Griffin, et al., Neutrophil-to-lymphocyte ratio is a predictive marker for invasive malignancy in intraductal papillary mucinous neoplasms of the pancreas, Ann. Surg. 2 (266) (2017) 339–345.
- [121] M.I. Canto, F. Harinck, R.H. Hruban, et al., International Cancer of the Pancreas Screening (CAPS) Consortium summit on the management of patients with increased risk for familial pancreatic cancer, Gut 3 (62) (2013) 339–347.
- [122] C.A. Barnes, E. Krzywda, S. Lahiff, et al., Development of a high risk pancreatic screening clinic using 3.0 T MRI, Fam. Cancer 1 (17) (2018) 101–111.
- [123] A. Gangi, M. MalafaandJ. Klapman, Endoscopic ultrasound-based pancreatic cancer screening of high-risk individuals: a prospective observational trial, Pancreas 5 (47) (2018) 586–591.
- [124] M.I. Canto, R.H. Hruban, E.K. Fishman, et al., Frequent detection of pancreatic lesions in asymptomatic high-risk individuals, Gastroenterology 4 (142) (2012) 796–804.
- [125] E. Ludwig, S.H. Olson, S. Bayuga, et al., Feasibility and yield of screening in relatives from familial pancreatic cancer families, Am. J. Gastroenterol. 5 (106) (2011) 946–954.
- [126] M.I. Canto, M. Goggins, R.H. Hruban, et al., Screening for early pancreatic neoplasia in high-risk individuals: a prospective controlled study, Clin. Gastroenterol. Hepatol. 6 (4) (2006) 766–781.
- [127] F. Harinck, I.C. Konings, I. Kluijt, et al., A multicentre comparative prospective blinded analysis of EUS and MRI for screening of pancreatic cancer in high-risk individuals, Gut 9 (65) (2016) 1505–1513.
- [128] M.T. Joergensen, A.M. Gerdes, J. Sorensen, et al., Is screening for pancreatic cancer in high-risk groups cost-effective? - experience from a Danish national screening program, Pancreatology 4 (16) (2016) 584–592.
- [129] H. Vasen, I. Ibrahim, C.G. Ponce, et al., Benefit of surveillance for pancreatic cancer in high-risk individuals: outcome of long-term prospective follow-up studies from three European expert centers, J. Clin. Oncol. 17 (34) (2016) 2010–2019.
- [130] M. Del Chiaro, C.S. Verbeke, N. Kartalis, et al., Short-term results of a magnetic resonance imaging-based Swedish screening program for individuals at risk for pancreatic cancer, JAMA Surg. 6 (150) (2015) 512–518.
- [131] W. Al-Sukhni, A. Borgida, H. Rothenmund, et al., Screening for pancreatic cancer in a high-risk cohort: an eight-year experience, J. Gastrointest. Surg. 4 (16) (2012) 771–783.
- [132] H.F. Vasen, M. Wasser, A. van Mil, et al., Magnetic resonance imaging surveillance detects early-stage pancreatic cancer in carriers of a p16-Leiden mutation, Gastroenterology 3 (140) (2011) 850–856.
- [133] P.A. Hartand, S.T. Chari, Is screening for pancreatic cancer in high-risk individuals one step closer or a fool's errand? Clin. Gastroenterol. Hepatol. 1 (17) (2019) 36–38.
- [134] Globocan, Survmark. https://gco.iarc.fr/survival/survmark/, 2018. (Accessed 10 January 2021). Accessed.
- [135] H. Zeng, W. Chen, R. Zheng, et al., Changing cancer survival in China during 2003-15: a pooled analysis of 17 population-based cancer registries. The Lancet, Glob. Health 5 (6) (2018) e555–e567.
- [136] P. Bailey, D.K. Chang, K. Nones, et al., Genomic analyses identify molecular subtypes of pancreatic cancer, Nature 7592 (531) (2016) 47–52.
- [137] E.A. Collisson, A. Sadanandam, P. Olson, et al., Subtypes of pancreatic ductal adenocarcinoma and their differing responses to therapy, Nat. Med. 4 (17) (2011) 500–503.
- [138] L. Buscail, B. BournetandP. Cordelier, Role of oncogenic KRAS in the diagnosis, prognosis and treatment of pancreatic cancer, Nat. Rev. Gastroenterol. Hepatol. 3 (17) (2020) 153–168.
- [139] H. Kim, B. Saka, S. Knight, et al., Having pancreatic cancer with tumoral loss of ATM and normal TP53 protein expression is associated with a poorer prognosis, Clin. Canc. Res. 7 (20) (2014) 1865–1872.
- [140] C. Song, T. Chen, L. He, et al., PRMT1 promotes pancreatic cancer growth and predicts poor prognosis, Cell. Oncol. 1 (43) (2020) 51–62.
- [141] W. Wang, W. Lou, B. Ding, et al., A novel mRNA-miRNA-lncRNA competing endogenous RNA triple sub-network associated with prognosis of pancreatic cancer, Aging 9 (11) (2019) 2610–2627.
- [142] S. Yokoyama, T. Hamada, M. Higashi, et al., Predicted prognosis of patients with pancreatic cancer by machine learning, Clin. Canc. Res. 10 (26) (2020) 2411–2421.